

6. Malcher, E. and Bodalski, T. (1969) *Diss. Pharm. Pharmacol.* **21**, 553.
7. Harborne, J. B. (1965) *Phytochemistry* **4**, 647.
8. Lamer, E., Malcher, E. and Grimshaw, J. (1968) *Tetrahedron Letters* **12**, 1419.
9. Torck, M. (1976) *Fitoterapia* **5**, 195.
10. Harborne, J. B., Mabry, T. J. and Mabry, H. (1975) *The Flavonoids*. Chapman & Hall, London.
11. Harborne, J. B. (1971) in *Chemotaxonomy of the Leguminosae* (Harborne, J. B., Boulter, D. and Turner, B. L., eds.) pp. 31–71. Academic Press, London.
12. Jay, M., Lebreton, Ph. and Letoublon, R. (1971) *Boissiera* **19**, 219.

Phytochemistry, Vol. 20, No. 8, pp. 2053–2054, 1981.
Printed in Great Britain.

0031-9422/81/082053-02 \$02.00/0
© 1981 Pergamon Press Ltd.

POLYPHENOLS FROM *ACHYROCLINE SATUREIODES**

GRACIELA E. FERRARO, CRISTINA NORBEDO and JORGE D. COUSSIO

Departamento de Bioquímica Vegetal, Cátedra de Farmacognosia, Facultad de Farmacia y Bioquímica, Universidad de Buenos Aires, Junin 956 (1113), Buenos Aires, Argentina

(Received 27 May 1980)

Key Word Index—*Achyrocline satureioides*; Compositae; galangin; galangin 3-methyl ether; quercetin 3-methyl ether; protocatheuoylcalleryanin; caffeoylcalleryanin.

Abstract—Galangin, galangin 3-methyl ether, quercetin, quercetin 3-methyl ether, caffeic acid and two esters of calleryanin (3,4-dihydroxybenzylalcohol 4-glucoside), with caffeic acid and protocathechuic acid, have been isolated from aerial parts of *Achyrocline satureioides*.

Achyrocline satureioides DC. (Lam.), which is distributed in dry regions of South America [1], is used in folk medicine [2–4]. Previously, Wagner *et al.* reported the isolation of isognaphaliin (3,7-dimethoxy-5,8-dihydroxyflavone) [5] and Ricciardi and Cassano described the components of the essential oil [6]. The present paper reports the isolation of galangin, galangin 3-methyl ether, quercetin, quercetin 3-methyl ether, caffeic acid and two esters of calleryanin (3,4-dihydroxybenzyl alcohol 4-glucoside) with caffeic acid and protocathechuic acid. Calleryanin was previously isolated from *Pyrus calleryana* [7] but this is the first report of its occurrence in the Compositae. Caffeic acid and its esters have been proved to increase bile flow in rats. The beneficial properties of *A. satureioides* are presumably related to the high content of caffeic acid esters [8].

EXPERIMENTAL

Plants were collected in Concepción del Uruguay, Province de Entre Rios, Argentina. Voucher specimens are deposited in the University Herbarium (Museo de Botánica, Universidad de Buenos Aires).

Extraction. Air-dried, ground aerial parts of *A. satureioides* (1.1 kg) were extracted with 50% aq. MeOH at room temp., the extracts evapd to dryness, taken into hot H₂O and partitioned with C₆H₆, CH₂Cl₂, Et₂O and EtOAc. The C₆H₆ extract was evapd to dryness and passed twice through a column packed with Sephadex LH20 and eluted with C₆H₆, CHCl₃ and MeOH. The CHCl₃ afforded 5,8-dihydroxy-3,7-dimethoxyflavone. The CHCl₃–MeOH eluates gave 5,7-dihydroxy-3-methoxyflavone (galangin 3-methyl ether). Spectral values and colour reactions for this compound were identical with previously reported values [9]. The Et₂O extract was evapd to dryness and passed through a column packed with polyamide. 3,5,7-Trihydroxyflavone (galangin) and quercetin crystallized from different fractions and were identified by mp, mmp, TLC and UV by comparison with authentic samples [9]. The EtOAc extract of *A. satureioides* was concd and run on 1D PC (Whatman No. 3) in

* Part 12 in the series "Flavonoids from Argentine Medicinal Plants". For Part 11 see Ferraro, G. E., Martino, V. S. and Coussio, J. D. (1977) *Phytochemistry* **16**, 1618.

H₂O, giving bands of caffeic acid, caffeoylcalleryanin and protocatchuoylcalleryanin which were eluted with MeOH and taken to dryness.

Caffeic acid (yield 0.5%) was crystallized from MeOH and determined by mp, mmp, TLC, PC and UV by comparison with an authentic sample [10].

Caffeoylcalleryanin (ester of caffeic acid with 3,4-dihydroxybenzyl alcohol 4-glucoside). Pale yellow amorphous powder. UV: pale blue; UV/NH₃: greenish grey. PC Whatman No. 1: 2% HOAc, H₂O, 0.1 N HCl and PhOH-H₂O (4:1) *R_f*s: 0.19, 0.10, 0.13 and 0.60, respectively. Found: C, 56.21; H, 5.22; C₂₂H₂₄O₁₁ requires: C, 56.89; H, 5.17%. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 244, 292 sh., 332; NaOMe: 267, 300 sh., 378 [7, 11].

Protocatechuoylcalleryanin (ester of protocatechuic acid with 3,4-benzyl alcohol 4-glucoside). Pale yellow amorphous powder. UV: very pale blue, UV/NH₃: yellow. Colour reactions: NH₃, yellow; FeCl₃, greenish grey. PC Whatman No. 1, 2% HOAc; H₂O; 0.1 N HCl and PhOH-H₂O (4:1) *R_f*s: 0.40, 0.48, 0.53 and 0.51, respectively. Found: C, 50.07; H, 5.62. C₂₀H₂₂O₁₁ · 2 H₂O requires: C, 50.63; H, 5.48%. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 260, 282 sh., 300; NaOMe: 240, 320.

Alkaline hydrolysis: Both caffeoylcalleryanin and protocatchuoylcalleryanin gave 3,6-dihydroxybenzyl alcohol 4-glucoside (calleryanin) on alkaline hydrolysis with 10 N KOH at room temp. for 30 min. Calleryanin was determined by the method of Challice and Williams [7]. Protocatechuic acid (3,4-dihydroxybenzoic acid) was determined by TLC, PC, UV spectroscopy and by comparison with an authentic sample. Caffeic acid (3,4-dihydroxycinnamic acid), extracted with Et₂O from the hydrolysate, was determined by the method of Nichiforescu-Coucou [10] and by UV, mp and mmp.

Enzymatic hydrolysis. Both caffeoylcalleryanin and protocatchuoylcalleryanin were hydrolysed with β -glucosidase, pH 5.0, at room temp. for 24 hr. Caffeic and protocatechuic acids and 3,4-dihydroxybenzyl alcohol were identified as above.

Glucose was determined by descending PC by comparison with an authentic sample. *

Acknowledgements—This work was supported in part by Consejo Nacional de Investigaciones Científicas y Técnicas 6324 d/78. We wish to thank Dr. Beatriz Sorarú, Facultad de Farmacia y Bioquímica, Universidad de Buenos Aires, for the collection and identification of plant material.

REFERENCES

1. Cabrera, A. (1963) *Flora de la Provincia de Buenos Aires*, p. 154. Colección Científica del INTA, Buenos Aires.
2. Parodi, D. (1979) *Enciclopedia Argentina de Agricultura y Jardinería*. ACME, Buenos Aires.
3. Paccard, E. (1905) *Lista de Algunas Plantas Medicinales de las Repúblicas Oriental y Argentina*. Montevideo.
4. Palma, N. H. (1973) *La Medicina Popular de la Puna*, p. 92, Cabargón, Buenos Aires.
5. Wagner, H., Maurer, G., Farkas, L., Hansel, R. and Ohlendorf, D. (1971) *Chem. Ber.* **104**, 2381.
6. Ricciardi, A. and Cassano, A. E. (1974) in *Catálogo Bibliográfico Fitoquímico Argentino III* (Amengual, B. M., ed.) Miscelánea No. 53. Tucumán, República Argentina.
7. Challice, J. S. and Williams, A. H. (1968) *Phytochemistry* **7**, 119.
8. Czook, G. and Schulze, P. J. (1973) *Z. Ernährungswiss* **12**, 224.
9. Mabry, T. J., Markham, K. R. and Thomas, M. B. (1970) *The Systematic Identification of Flavonoids*. Springer, Berlin.
10. Martino, V. S., Debenedetti, S. L. and Coussio, J. D. (1979) *Phytochemistry* **18**, 2052.
11. Herrman, Von K. (1978) in *Progress in the Chemistry of Organic Natural Products* (Herz, W., Grisebach, H. and Kirby, G. W., eds.) p. 73. Springer, Wien.